Vitamin D Regulated Keratinocyte Differentiation: Role of Coactivators

D.D. Bikle,* C.-L. Tu, Z. Xie, and Y. Oda

Departments of Medicine and Dermatology, Veterans Affairs Medical Center and University of California, San Francisco, CA 94121

Abstract 1,25 Dihydroxyvitamin D (1,25(OH)₂D) regulates the differentiation of keratinocytes. 1,25(OH)₂D raises intracellular free calcium (Cai) as a necessary early step toward stimulating differentiation. $1,25(OH)₂D$ induces the calcium sensing receptor (CaR) in keratinocytes and enhances the calcium response of these cells. Activation of the CaR by calcium increases intracellular free calcium by a mechanism involving phospholipase C (PLC) cleavage of phosphatidylinositolbisphosphate into inositoltrisphosphate (IP₃) and diacylglycerol (DG). 1,25(OH)₂D induces the family of PLCs. PLC-g1 has a DR6 VDRE in its promoter which binds and is activated by VDR/RAR rather than VDR/RXR. The involucrin gene, which encodes a critical component of the cornified envelope, contains a DR3 VDRE in its promoter that acts in conjunction with a nearby AP-1 site. The sequential regulation of these genes is critical for the differentiation process. In undifferentiated keratinocytes, the VDR binds preferentially to the DRIP complex of coactivators. However, with differentiation DRIP 205 is no longer produced, and the VDR switches partners to the SRC family (SRC2 and 3). These studies suggest that at least part of the sequential activation of genes required during keratinocyte differentiation is regulated by the change (availability) of these different coactivator complexes. J. Cell. Biochem. 88: 290–295, 2003. © 2002 Wiley-Liss, Inc.

Key words: involucrin; transglutaminase; vitamin D receptor; steroid receptor coactivator; phospholipase C; calcium receptor

The observation that $1,25(OH)_2D$ induces keratinocyte differentiation was first made by Hosomi et al. [1983] and provided a rationale for the previous and unexpected finding of $1,25(OH)₂D$ receptors in the epidermis [Stumpf] et al., 1979]. $1,25(OH)₂D$ is likely to be an autocrine or paracrine factor for epidermal differentiation since it is produced by the keratinocyte, but under normal circumstances keratinocyte production of $1,25(OH)_2D$ does not appear to contribute to circulating levels [Bikle

*Correspondence to: D.D. Bikle, MD, PhD, Veterans Affairs Medical Center (111N), 4150 Clement St., San Francisco, CA 94121. E-mail: doctor@itsa.ucsf.edu

Received 20 June 2002; Accepted 24 June 2002

DOI 10.1002/jcb.10339

et al., 1986]. Recent evidence that the keratinocyte also converts vitamin D to 25OHD indicates that the keratinocyte is unique in its possession of the complete pathway from 7 dehydrocholesterol to 1,25(OH)2D [Lehmann et al., 1999]. The receptors for and the production of $1,25(OH)_2D$ vary with differentiation [Pillai et al., 1988] in a manner which suggests feedback regulation; both are reduced in the later stages of differentiation. The differentiation pathway that we have studied most directly is the formation of the cornified envelope (CE). $1,25(OH)_2D$ increases gene expression and the mRNA and protein levels for involucrin, a precursor for the CE, and transglutaminase, the enzyme which crosslinks involucrin and other substrates to form the CE. Thus, $1,25(OH)_{2}D$ promotes CE formation at subnanomolar concentrations in preconfluent keratinocytes [Hosomi et al., 1983; Smith et al., 1986; McLane et al., 1990; Pillai and Bikle, 1991; Su et al., 1994]. At these same concentrations, $1,25(OH)_2D$ may either promote or inhibit proliferation [Itin et al., 1994; Bollag et al., 1995; Gniadecki, 1996] indicating

Grant sponsor: Department of Veterans Affairs; Grant sponsor: National Institutes of Health; Grant numbers: RO1 AR38386, PO1 AR39448; Grant sponsor: American Institute for Cancer Research; Grant number: 98A079.

2002 Wiley-Liss, Inc.

that differentiation is not dependent on the antiproliferative actions of $1,25(OH)_2D$. Concentrations of $1,25(OH)_2D$ above $10^{-9}M$ are generally found to have only an antiproliferative effect. The mechanisms underlying the proproliferative actions are not known. The antiproliferative effects are accompanied by an increase in TGF- β [Kim et al., 1992] and a reduction in c-myc and the EGF receptor mRNA levels [Matsumoto et al., 1990]. In other cell lines, the antiproliferative actions are associated with an arrest at G1 with an increase in the inhibitors of G1 progression including p21waf1, p27kip1, and ink4 family members (p15, p16, p18) [Liu et al., 1996], and a decrease in the phosphorylation of the retinoblastoma protein pRB.

INTERACTION OF 1,25(OH)₂D AND CALCIUM

Calcium clearly affects the ability of $1,25(OH)₂D$ to stimulate keratinocyte differentiation, and vice versa. We [Su et al., 1994] observed that both calcium in the absence of $1,25(OH)_{2}D$ and $1,25(OH)_{2}D$ at low (0.03 mM) calcium raise the mRNA levels for involucrin and transglutaminase in a dose dependent fashion by stimulating gene expression. The stimulation of mRNA levels by calcium and $1,25(OH)₂D$ is synergistic at early time points, 4 h after addition of calcium and 28 h after addition of $1,25(OH)₂D$, but longer periods of incubation lead to a paradoxical fall in the mRNA levels for these proteins. This is due to the fact that although transcription is increased by calcium and $1,25(OH)_2D$, stability of the mRNA is reduced in cells incubated with calcium and $1,25(OH)₂D$ also in a synergistic fashion. The mechanism by which mRNA stability is regulated by calcium and $1,25(OH)₂D$ remains unexplored.

1,25(OH)2D RAISES CAI BY INDUCING THE CALCIUM RECEPTOR

The ability of $1,25(OH)_2D$ to increase intracellular free calcium levels (Cai) [Pillai and Bikle, 1991] accounts for at least part of the ability of $1,25(OH)₂D$ to induce differentiation. The keratinocyte responds to extracellular calcium via the same calcium receptor (CaR) found in the parathyroid gland and other tissues [Bikle et al., 1996; Oda et al., 1998; Tu et al., 1999]. This CaR is critical for calcium induced differentiation [Oda et al., 2000; Tu et al., 2001] in that knocking out the CaR in mice [Ho et al., 1995; Oda et al., 2000] or in human keratinocytes by antisense constructs [Tu et al., 2001] leads to keratinocytes which fail to respond to calcium with respect to increases in Cai and/or increases in the differentiation markers. $1,25(OH)_2D$ increases the mRNA levels of the CaR and enhances the responsiveness of the keratinocyte to calcium [Ratnam et al., 1999]. One interesting twist in this story is that keratinocytes make both a full length form of the CaR and an alternatively spliced form in which exon 5 is spliced out [Oda et al., 1998]. The alteratively spliced CaR does not mediate the acute response to calcium (by increases in either Cai or IP_3), and its function is unclear [Oda et al., 1998; Oda et al., 2000].

1,25(OH)2D RAISES CAI BY INDUCING THE PHOSPHOLIPASE C (PLC) FAMILY

A second mechanism by which $1,25(OH)₂D$ increases Cai involves the induction of the family of PLCs [Pillai et al., 1995]. As a result, $1,25(OH)₂D$ was found to increase both the basal and hormone (calcium, ATP, bradykinin, histamine) stimulated levels of IP_3 and Cai [Pillai et al., 1995]. We [Xie and Bikle, 1997] then investigated the mechanism by which $1,25(OH)₂D$ induced the PLC family, focusing on PLC- γ 1 because it is the most abundant PLC in keratinocytes [Punnonen et al., 1993] and potentially the most important with respect to mediating the effects of $1,25(OH)_2D$ and calcium on proliferation and differentiation. The $5'$ -flanking region of the human PLC- γ 1 gene was isolated from a human P1 genomic DNA library. Deletion and mutation studies of this fragment demonstrated a VDRE that contains a motif arranged as two direct repeats separated by six bases (DR6) between -786 and -803 base pairs (Fig. 1). Incubation of the oligonucleotide containing the DR6 with keratinocyte nuclear extracts produced a specific protein–DNA complex that shifted to a higher molecular weight form upon the addition of an antibody to the vitamin D receptor (VDR) and to the retinoic acid receptor (RAR) but not to the retinoid X receptor (RXR). These results indicate that for this VDRE, the VDR couples to RAR [Xie and Bikle, 1998]. All trans retinoic acid (tRA) potentiated the ability of $1,25(OH)_2D$ to stimulate not only the PLC- γ 1 VDRE/luciferase

Fig. 1. The sequence of the vitamin D response element (VDRE) in the phospholipase $C-\gamma$ 1 promoter. Unlike most VDREs, this VDRE has six nucleotides between the two half sites (DR6). This VDRE is also unusual in that it recognizes VDR/RAR heterodimers rather than VDR/RXR heterodimers.

construct in transfected keratinocytes but also the mRNA levels for PLC- γ 1 in the normal keratinocytes [Xie and Bikle, 1998].

Although the induction of $PLC-\gamma1$ by 1,25(OH)2D could explain part of the ability of $1,25(OH)₂D$ to raise Cai, it was not clear how important this mechanism was to $1,25(OH)_2D$ regulated differentiation. Accordingly, we [Xie and Bikle, 1999] blocked expression of PLC- γ 1 in human keratinocytes by transfecting cells with an antisense human PLC- γ 1 cDNA construct. These cells demonstrated a specific reduction in PLC- γ 1 protein levels compared to the empty vector-transfected cells and a marked reduction in the mRNA and protein levels of the differentiation markers, involucrin and transglutaminase, following administration of $1,25(OH)₂D$ [Xie and Bikle, 2001].

INVOLUCRIN PROMOTER VITAMIN D RESPONSE ELEMENT

We [Ng et al., 2000] mapped the calcium responsive region of the involucrin promoter to a region \sim 2,100 bases 5' of the translation start site, and called this the calcium response element (CaRE) (Fig. 2). This region contains an AP-1 site which is critical for basal as well as calcium and $1,25(OH)₂D$ stimulation of involucrin gene expression in that mutations of this AP-1 site lower basal expression and block calcium and $1,25(OH)₂D$ stimulation. Separated from this AP-1 site by an SP-1 site is a vitamin D response element (VDRE) with a DR3 configuration. This VDRE binds VDR/RXR complexes. Mutation of the VDRE blocks $1,25(OH)₂D$ stimulation of involucrin gene expression, but does not alter basal activity or calcium stimulation [Bikle et al., 2002].

Fig. 2. The sequence of the calcium response element (CaRE) in the involucrin promoter. The CaRE contains an AP-1 site which is essential for basal activity as well as the ability of both calcium and $1,25(OH)_{2}D$ to stimulate involucrin gene expression. In contrast, the DR3 appears to mediate only $1,25(OH)_2D$ regulated expression of the involucrin gene. The functions of the SP-1 site are less clearly established.

SEQUENTIAL ACTIVATION OF THE VITAMIN D RECEPTOR BY DRIP AND SRC COACTIVATOR COMPLEXES

The control of keratinocyte proliferation and differentiation by $1,25(OH)₂D$ appears to involve a sequential turning on and off of genes as the cells progress from the proliferating basal cell to the fully differentiated corneocyte. As a first step to determine whether this transition involved a change in the regulation of the VDR function itself, we [Oda et al., submitted] examined the ligand dependent binding of proteins to the VDR at different stages of differentiation. A GST-VDR construct (gift from Dr. Paul McDonald) was expressed and used to bind proteins from nuclear extracts of undifferentiated and differentiated (by calcium) keratinocytes. The bound proteins were eluted from the GST-VDR column, separated by SDS-PAGE, and the proteins identified by mass spectroscopy. The complex from the proliferating cells was nearly identical to that previously identified in other cells by other groups and named vitamin D receptor interacting proteins (DRIP) [Rachez et al., 1999], thyroid receptor activating proteins (TRAP) [Yuan et al., 1998], or activator receptor complex (ARC) [Naar et al., 1999]. When nuclear extracts were obtained from differentiated keratinocytes, the complex was quite different. The major anchoring protein of the DRIP complex, namely DRIP205, was no longer present. Instead, two members of the steroid receptor coactivator (SRC) family were present, SRC 2 and 3, along with several smaller members of the DRIP complex. Western analysis of cellular extracts from keratinocytes at different stages of differentiation showed a decrease in DRIP 205 with differentiation, but no change in SRC 2 or 3. Transfection of undifferentiated keratinocytes with DRIP205 or SRC 3 potentiated the ability of $1,25(OH)_2D$ to stimulate expression of a VDRE/luciferase construct. However, when the same experiment was performed in differentiated keratinocytes, only transfection with SRC3 enhanced the response to $1,25(OH)₂D$. These results suggest that DRIP may be more important in mediating the effects of $1,25(OH)_2D$ on the earliest stages of differentiation, but SRC 2/3 may be more important during the latter stages of differentiation (Fig. 3). We are currently testing this hypothesis.

INTEGRATION MODEL

Our current working model for the mechanisms by which $1,25(OH)₂D$ regulates keratinocyte differentiation is shown in Figure 4. The

Fig. 3. Model showing selective utilization of two distinct VDR coactivators, DRIP/mediator and SRC/p160, in transcriptional activation during keratinocyte differentiation. Both DRIP/mediator and SRC/p160 may be involved in vitamin D regulated transcription of undifferentiated proliferating cells (upper). At this stage, transcriptional activation may require primarily the linking of the VDR complex to the general transcription machinery including RNA Pol II. After the cells are differentiated, DRIP205 apparently is no longer required, and SRC/p160 family members take the dominant role in transcriptional control (lower).

Fig. 4. A model of the interactions between calcium and $1,25(OH)₂D$ in regulating keratinocyte differentiation. The essential features of this model include the G protein coupled calcium sensing receptor (CaR) which when activated by calcium leads to the activation of phospholipase C (PLC), thus producing two important second messengers from the hydrolysis of phosphatidylinositol bisphosphate (PIP2)—diacyl glycerol (DAG) and inositol tris phosphate (IP₃). IP₃ stimulates the release of calcium from intracellular stores, thus increasing intracellular free calcium concentrations (Cai). The increase in DAG and Cai activate protein kinases (PKC). PKC in turn induces and/or activates the AP-1 family of transcription factors which regulate the expression of the genes encoding the proteins such as involucrin, transglutaminase (TG) and loricrin involved in cornified envelope (CE) formation. The increase in Cai may also stimulate the influx of calcium through calcium channels thus prolonging the Cai signal. $1,25(OH)_2D$ increases the expression of both the CaR and PLC, enhancing the cellular response to calcium. Both calcium and $1,25(OH)_{2}D$ may regulate the transcription of involucrin, TG, and loricrin by mechanisms other than through AP-1.

keratinocyte expresses a CaR which by coupling to and activating PLC controls the production of two important second messengers, $IP₃$ and DAG. IP₃ stimulates the release of calcium from intracellular stores thus raising Cai. The rise in DAG and Cai activate PKC. The initial rise in Cai may also trigger the influx of calcium through calcium channels, maintaining an elevation in Cai for a prolonged period of time. Activated PKC leads to the induction and activation of AP-1 transcription factors which regulate the transcription of a number of genes including keratin 1, transglutaminase, involucrin, loricrin, and profilaggrin which are required for the differentiation process. $1,25(OH)_{2}D$, which is produced by the keratinocyte in a highly regulated fashion, modulates calcium regulated differentiation at several steps. First, $1,25(OH)₂D$ increases CaR expression, thus making the cell more responsive to calcium. Secondly, $1,25(OH)_2D$ induces all the PLCs again increasing the responsiveness of the cell to calcium. Finally, $1,25(OH)_2D$ may have a direct effect on the transcription of the genes such as involucrin in addition to promoting their expression by increasing Cai. Differentiation requires a carefully orchestrated turning on and off of genes critical for the differentiation process. Sequential activation of the VDR by the DRIP and SRC coactivator complexes may provide such control.

ACKNOWLEDGMENTS

The authors acknowledge the administrative support of Vivian Wu and the financial support from the Department of Veterans Affairs, the National Institutes of Health (RO1 AR38386, PO1 AR39448), and the American Institute for Cancer Research (98A079).

REFERENCES

- Bikle DD, Nemanic MK, Whitney JO, Elias PW. 1986. Neonatal human foreskin keratinocytes produce 1,25 dihydroxyvitamin D3. Biochemistry 25:1545–1548.
- Bikle DD RA, Mauro T, Harris J, Pillai S. 1996. Changes in calcium responsiveness and handling during keratinocyte differentiation: potential role of the calcium receptor. J Clin Invest 97:1085–1093.
- Bikle DD, Ng D, Oda Y, Hanley K, Feingold K, Xie Z. 2002. The vitamin D response element of the involucrin gene mediates its regulation by 1,25 dihydroxyvitamin D. J Invest Dermatol (in press).
- Bollag WB, Ducote J, Harmon CS. 1995. Biphasic effect of 1,25-dihydroxyvitamin D3 on primary mouse epidermal keratinocyte proliferation. J Cell Physiol 163:248–256.
- Gniadecki R. 1996. Stimulation versus inhibition of keratinocyte growth by 1,25-Dihydroxyvitamin D3: dependence on cell culture conditions. J Invest Dermatol 106:510–516.
- Ho C, Conner DA, Pollak MR, Ladd DJ, Kifor O, Warren HB, Brown EM, Seidman JG, Seidman CE. 1995. A mouse model of human familial hypocalciuric hypercalcemia and noonatal severe hyperparathyroidism. Nature Genet 11:389–394.
- Hosomi J, Hosoi J, Abe E, Suda T, Kuroki T. 1983. Regulation of terminal differentiation of cultured mouse epidermal cells by 1 alpha,25-dihydroxyvitamin D3. Endocrinology 113:1950–1957.
- Itin PH, Pittelkow MR, Kumar R. 1994. Effects of vitamin D metabolites on proliferation and differentiation of cultured human epidermal keratinocytes grown in serumfree or defined culture medium. Endocrinology 135: 1793–1798.
- Kim HJ, Abdelkader N, Katz M, McLane JA. 1992. 1,25- Dihydroxy-vitamin-D3 enhances antiproliferative effect and transcription of TGF-betal on human keratinocytes in culture. J Cell Physiol 151:579–587.
- Lehmann B, Tiebel O, Meurer M. 1999. Expression of vitamin D3 25-hydroxylase (CYP27) mRNA after induction by vitamin D3 or UVB radiation in keratinocytes of

human skin equivalents–a preliminary study. Arch Dermatol Res 291:507–510.

- Liu M, Lee MH, Cohen M, Bommakanti M, Freedman LP. 1996. Transcriptional activation of the Cdk inhibitor p21 by vitamin D3 leads to the induced differentiation of the myelomonocytic cell line U937. Genes Dev 10:142–153.
- Matsumoto K, Hashimoto K, Nishida Y, Hashiro M, Yoshikawa K. 1990. Growth-inhibitory effects of 1,25 dihydroxyvitamin D3 on normal human keratinocytes cultured in serum-free medium. Biochem Biophys Res Commun 166:916–923.
- McLane JA, Katz M, Abdelkader N. 1990. Effect of 1,25 dihydroxyvitamin D3 on human keratinocytes grown under different culture conditions. In Vitro Cell Dev Biol 26:379–387.
- Naar AM, Beaurang PA, Zhou S, Abraham S, Solomon W, Tjian R. 1999. Composite co-activator ARC mediates chromatin-directed transcription activation. Nature 398: 828–833.
- Ng DC SS, Ree D, Bikle DD. 2000. Requirement of an AP-1 site in the calcium response region of the involucrin promoter. J Biol Chem 275:24080–24088.
- Oda Y, Tu CL, Pillai S, Bikle DD. 1998. The calcium sensing receptor and its alternatively spliced form in keratinocyte differentiation. J Biol Chem 273:23344–23352.
- Oda Y, Tu CL, Chang W, Crumrine D, Kömüves L, Mauro T, Elias PM, Bikle DD. 2000. The calcium sensing receptor and its alternatively spliced form in murine epidermal differentiation. J Biol Chem 275:1183–1190.
- Pillai S, Bikle DD. 1991. Role of intracellular-free calcium in the cornified envelope formation of keratinocytes: differences in the mode of action of extracellular calcium and 1,25 dihydroxyvitamin D3. J Cell Physiol 146:94– 100.
- Pillai S, Bikle DD, Elias PM. 1988. 1,25-Dihydroxyvitamin D production and receptor binding in human keratinocytes varies with differentiation. J Biol Chem 263:5390– 5395.
- Pillai S, Bikle DD, Su MJ, Ratnam A, Abe J. 1995. 1,25- Dihydroxyvitamin D3 upregulates the phosphatidylinositol signaling pathway in human keratinocytes by increasing phospholipase C levels. J Clin Invest 96: 602–609.
- Punnonen K, Denning M, Lee E, Li L, Rhee SG, Yuspa SH. 1993. Keratinocyte differentiation is associated with changes in the expression and regulation of phospholipase C isoenzymes. J Invest Dermatol 101:719–726.
- Rachez C, Lemon BD, Suidan Z, Bromleigh V, Gamble M, Naar AM, Erdjument-Bromage H, Tempst P, Freedman LP. 1999. Ligand-dependent transcription activation by nuclear receptors requires the DRIP complex. Nature 398:824–828.
- Ratnam AV, Bikle DD, Cho JK. 1999. 1,25 dihydroxyvitamin D3 enhances the calcium response of keratinocytes. J Cell Physiol 178:188–196.
- Smith EL, Walworth NC, Holick MF. 1986. Effect of 1 alpha,25-dihydroxyvitamin D3 on the morphologic and biochemical differentiation of cultured human epidermal keratinocytes grown in serum-free conditions. J Invest Dermatol 86:709–714.
- Stumpf WE, Sar M, Reid FA, Tanaka Y, DeLuca HF. 1979. Target cells for 1,25-dihydroxyvitamin D3 in intestinal tract, stomach, kidney, skin, pituitary, and parathyroid. Science 206:1188–1190.
- Su M-J, Bikle DD, Mancianti ML, Pillai S. 1994. 1,25-Dihydroxyvitamin D3 potentiates the keratinocyte response to calcium. J Biol Chem 269:14723– 14729.
- Tu C-L, Oda Y, Bikle DD. 1999. Effects of calcium receptor activator on the cellular response to calcium in human keratinocytes. J. Invest Dermatol 113:340–345.
- Tu C-L, Chang W, Bikle DD. 2001. The extracellular calcium-sensing receptor is required for calcium-induced differentiation in human keratinocytes. J Biol Chem 276: 41079–41085.
- Xie Z, Bikle DD. 1997. Cloning of the human phospholipase $C-\gamma 1$ promoter and identification of a DR-6 type vitamin D responsive element. J Biol Chem 272:6573– 6577.
- Xie Z, Bikle DD. 1998. Differential regulation of vitamin D responsive elements in normal and transformed keratinocytes. J Invest Dermatol 110:730–733.
- Xie Z, Bikle DD. 1999. Phospholipase C-gamma1 is required for calcium-induced keratinocyte differentiation. J Biol Chem 274:20421–20424.
- Xie Z, Bikle DD. 2001. Inhibition of 1,25-dihydroxyvitamin D-induced keratinocyte differentiation by blocking the expression of phospholipase C- γ 1. J Invest Dermatol 117: 1250–1254.
- Yuan C, Ito M, Fondell JD, Fu ZY, Roeder RG. 1998. The TRAP220 component of a thyroid hormone receptorassociated protein (TRAP) coactivator complex interacts directly with nuclear receptors in a ligand-dependent fashion. Proc Natl Acad Sci USA 95:7939–7944.